

Test Specific Guidelines

Hereditary Connective Tissue Disorder Testing

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Procedures Addressed

The inclusion of any procedure code in this table is provided for informational purposes and is not a guarantee of coverage nor an indication that prior authorization is required.

<u>Procedures addressed by this guideline</u>	<u>Procedure codes</u>
<u>Aortic Dysfunction or Dilation Duplication/Deletion Analysis Panel</u>	<u>81411</u>
<u>Aortic Dysfunction or Dilation Genomic Sequencing Analysis Panel</u>	<u>81410</u>
<u>Hereditary Connective Tissue Disorder Gene Analysis</u>	<u>81400</u> <u>81401</u> <u>81402</u> <u>81403</u> <u>81404</u> <u>81405</u> <u>81406</u> <u>81407</u> <u>81408</u> <u>81479</u>
<u>Hereditary Connective Tissue Disorder Known Familial Mutation Analysis</u>	<u>81403</u>

What Are Hereditary Connective Tissue Disorders?

Definition

Hereditary connective tissue disorders (HCTDs) are a group of disorders that

affect the connective tissues that support the skin, bones, joints, heart, blood vessels, eyes, and other organs.¹

While specific features vary by type, an unusually large range of joint movement (hypermobility) and cardiovascular disease (such as thoracic aortic aneurysms and dissections, or TAAD) are features that are present in many HCTDs. Medical management may differ based on the underlying genetic etiology.

In many cases, a careful clinical examination by a specialist familiar with clinical features of these conditions can help to point toward one condition or group of conditions. In these cases, testing for gene(s) associated with a single condition or group of conditions would be most appropriate. However, in some cases, it can be difficult to reliably diagnose an HCTD based on clinical and family history alone.

Although connective tissue disorders as a whole are common, individual hereditary connective tissue disorders are relatively uncommon.¹

There are more than 200 connective tissue disorders.² Some of the most common types are summarized below:

Arterial tortuosity syndrome (ATS) — An autosomal recessive disorder associated with severe and widespread tortuosity of the aorta and middle-sized arteries, with an increased risk of aneurysms and dissections. Other features include stenosis of the aorta and/or pulmonary arteries, characteristic facies with high palate and dental crowding, and soft/doughy skin. Additional connective tissue disorder features that may be present include skeletal findings (scoliosis, pectus anomalies, joint laxity), hernias, hypotonia, and ocular involvement (myopia, keratoconus). SLC2A10 is the only gene known to be associated with ATS. Sequence variants are the most common; exon deletions have been reported in a couple cases.³

Congenital contractural arachnodactyly (Beals syndrome) — An autosomal dominant disorder characterized by a Marfan-like appearance (tall, slender habitus in which arm span exceeds height) and long, slender fingers and toes (arachnodactyly). Most affected individuals have a “crumpled” appearance to their ears and most have contractures of major joints (knees and ankles) at birth. Hip contractures, adducted thumbs, and club foot may occur. The majority of affected individuals have muscular hypoplasia. Kyphosis/scoliosis is present in about half of all affected individuals. Dilatation of the aorta is occasionally present. “FBN2 is the only gene in which mutation is known to cause congenital contractural arachnodactyly.”⁴

Cutis laxa — A group of disorders characterized by lax, sagging skin that often hangs in loose folds, causing the face and other parts of the body to have a droopy appearance. Extremely wrinkled skin may be particularly noticeable on the neck and in the armpits and groin. Other features may include arterial aneurysm and dissection, emphysema, and inguinal or umbilical hernia. There are autosomal dominant, autosomal recessive, and X-linked forms. Causative

autosomal genes include ELN, FBLN5, ATP6V0A2, EFEMP2, and LTBP4.^{5,6} The X-linked form is due to mutations in ATP7A (see also Occipital Horn Syndrome).⁵

Ehlers Danlos syndromes (EDS) — A heterogeneous group of disorders, the majority of which share the features of joint hypermobility and skin involvement. There are 13 types: classical, classical-like, cardiac-valvular, vascular, hypermobile (includes “joint hypermobility syndrome”), arthrochalasia, dermatosparaxis, kyphoscoliotic, spondylodysplastic, musculocontractural, myopathic, periodontal, and brittle cornea syndrome. Some types have autosomal dominant inheritance, while others are autosomal recessive. Hypermobile type is the most common, but its genetic etiology is currently unknown. Genetic testing is available for the other EDS types (see Table 1 below for a list of genes).^{7,8}

Homocystinuria due to cystathionine beta-synthase deficiency — An autosomal recessive metabolic disorder in which affected individuals have markedly elevated plasma total homocysteine and methionine. Clinical features include involvement of the eye (ectopia lentis and/or severe myopia), skeletal system (excessive height, long limbs, scoliosis, and pectus excavatum), and vascular system (thromboembolism). Many have developmental delay/intellectual disability. Treatment involves maintenance of normal or near-normal plasma homocysteine concentrations using a specialized diet and vitamin supplementation. The diagnosis can be substantiated by detection of biallelic pathogenic mutations in the CBS gene. Sequence analysis detects 95-98% of mutations, while deletion/duplication analysis detects <5%.⁹

Loeys-Dietz syndrome (LDS) — LDS is an autosomal dominant disorder that affects many parts of the body.¹⁰ LDS is caused by mutations in six genes: TGFB2 (55-60%), TGFB1 (20-25%), SMAD3 (5-10%), TGFB2 (5-10%), TGFB3 (1-5%), or SMAD2 (1-5%). Major manifestations of this condition include “vascular findings (dilatation or dissection of the aorta, other arterial aneurysms or tortuosity), skeletal findings (pectus excavatum or pectus carinatum, scoliosis, joint laxity or contracture, long thin fingers and toes, cervical spine malformation and/or instability), craniofacial findings (widely spaced eyes, bifid uvula/cleft palate, craniosynostosis), and cutaneous findings (translucent skin, easy bruising, dystrophic scars).”¹⁰ Given that there is no clinical diagnostic criteria established for LDS, genetic testing, either through serial single-gene testing or use of a multigene panel, can establish the diagnosis.¹⁰

Marfan syndrome (MFS) — MFS is an autosomal dominant disorder that affects connective tissue in many parts of the body.¹¹ MFS is caused by mutations in the FBN1 gene. Up to 93% of people meeting diagnostic criteria for MFS will have a mutation in this gene. Diagnostic criteria, called the Ghent criteria, exists for MFS. Major manifestations of the disease include aortic enlargement and ectopia lentis. Other features include, but are not limited to, bone overgrowth and joint laxity, long arms and legs, scoliosis, sternum deformity (pectus excavatum or carinatum), long thin fingers and toes, dural ectasia (stretching of the dural sac),

hernias, stretch marks on the skin, and lung bullae. Symptoms can present in males or females at any age. Symptoms typically worsen over time. Infants who present with symptoms typically have the most severe disease course.¹¹

NOTCH1-related aortic valve disease — NOTCH1 variants can be associated with autosomal dominant congenital heart defects affecting the left ventricular outflow tract (LVOT), most commonly bicuspid aortic valve (BAV). Adult-onset aortic valve calcification is a frequent feature. NOTCH1 variants have also been identified in 4.2% of individuals with sporadic BAV and much less frequently with other LVOT malformations. Mutations in this gene are also associated with Adams-Oliver syndrome, which is characterized by aplasia cutis congenita of the scalp and malformations of the limbs, brain, and cardiovascular system.¹²

Osteogenesis imperfecta (OI) — A group of disorders associated with a propensity to fractures with little or no trauma. Additional features may include skeletal anomalies, short stature, hearing loss, and blue/gray sclera. The severity is highly variable, ranging from a mild form with few fractures and normal life expectancy, to severe forms with neonatal lethality. OI types I-IV account for the majority of cases, and are caused by heterozygous mutations in the COL1A1 and COL1A2 genes. Inheritance is autosomal dominant. Autosomal recessive forms of OI are rare, and can be associated with mutations in a number of different genes.¹³

FLNA Deficiency — FLNA deficiency is associated with a phenotypic spectrum that includes FLNA-related periventricular nodular heterotopia (PVNH). FLNA deficiency is an X-linked condition that is prenatally or neonatally lethal in most males. Therefore, most affected individuals are female. In addition to PVNH, some individuals have connective tissue anomalies such as joint hypermobility, aortic dilation, and other vascular anomalies. 93% of individuals with FLNA-related PVNH have a sequence variant; genomic rearrangements have been reported in a few cases.¹⁴

Stickler syndrome — A disorder characterized by ocular findings (myopia, cataract and retinal detachment), hearing loss, craniofacial findings (midfacial underdevelopment and cleft palate), mild spondyloepiphyseal dysplasia and/or early-onset arthritis. Clinical diagnostic criteria are available. >90% of cases are due to mutations in COL2A1 or COL11A1. Mutations in these genes, and COL11A2, are inherited in an autosomal dominant pattern. Mutations in COL9A1, COL9A2, and COL9A3 are rare, and inherited in an autosomal recessive pattern.¹⁵

Thoracic Aortic Aneurysm and Dissection (TAAD) — Familial TAAD is defined as dilatation and/or dissection of the thoracic aorta, absence of clinical features of MFS, LDS or vascular EDS, and a positive family history of TAAD. Approximately 30% of families with heritable thoracic aortic disease (HTAD) who do not have a clinical diagnosis of MFS or another syndrome have a causative mutation in one of 14 known HTAD-related genes (see Table 1 below).¹⁶

Test Information

Introduction

Testing for hereditary connective tissue disorders may include next-generation sequencing or multigene panels.

Next Generation Sequencing Assay

Next generation sequencing (NGS), which is also sometimes called massively parallel sequencing, was developed in 2005 to allow larger scale and more efficient gene sequencing. NGS relies on sequencing many copies of small pieces of DNA simultaneously and using bioinformatics to assemble the sequence. Sequence analysis detects single nucleotide substitutions and small (several nucleotide) deletions and insertions. Regions analyzed typically include the coding sequence and intron/exon boundaries. Promoter regions and intronic sequences may also be sequenced if disease-causing mutations are known to occur in these regions of a gene.

Multi-Gene Testing Panels

The efficiency of NGS has led to an increasing number of large, multi-gene testing panels. NGS panels that test several genes at once are particularly well-suited to conditions caused by more than one gene or where there is considerable clinical overlap between conditions making it difficult to reliably narrow down likely causes. Additionally, tests should be chosen to maximize the likelihood of identifying mutations in the genes of interest, contribute to alterations in patient management, and/or minimize the chance of finding variants of uncertain clinical significance.

Clinical genetic testing is available for many HCTDs. However, hypermobile EDS (hEDS), joint hypermobility syndrome, and isolated joint hypermobility, including “hypermobility spectrum disorders”, continue to require a clinical diagnosis, since the genetic etiology of these disorders is not yet known.⁸

Guidelines and Evidence

No current U.S guidelines address the use of multi-gene panels in HCTDs.

An expert-authored review (updated in 2018)¹⁷ states the following regarding hEDS: “If an individual's personal or family history is suggestive of one of the other types of EDS or another hereditary disorder of connective tissue or arterial fragility syndrome..., analysis of an associated gene or multi-gene connective tissue disease panel may be appropriate. Failure to identify a pathogenic variant with such multiple gene testing reduces the likelihood of an arterial fragility syndrome, but does not completely rule it out, especially in the setting of a positive personal or family history of arterial fragility. Negative testing for an arterial fragility syndrome also does not confirm a diagnosis of hEDS. Therefore,

such testing is not recommended in the absence of specific suggestive signs, symptoms, or family history.”

According to the International Consortium on the Ehlers-Danlos Syndromes (2017):⁸

“In view of the vast genetic heterogeneity and phenotypic variability of the EDS subtypes, and the clinical overlap between many of these subtypes, but also with other HCTDs, the definite diagnosis relies for all subtypes, except hEDS, on molecular confirmation with identification of (a) causative variant(s) in the respective gene.”

“Molecular diagnostic strategies should rely on NGS technologies, which offer the potential for parallel sequencing of multiple genes. Targeted resequencing of a panel of genes...is a time- and cost-effective approach for the molecular diagnosis of the genetically heterogeneous EDS. When no mutation (or in case of an autosomal recessive condition only one mutation) is identified, this approach should be complemented with a copy number variant (CNV) detection strategy to identify large deletions or duplications, for example Multiplex Ligation-dependent Probe Amplification (MLPA), qPCR, or targeted array analysis.”

“The diagnosis of hEDS remains clinical as there is yet no reliable or appreciable genetic etiology to test for in the vast majority of patients.”

Criteria

This guideline applies to hereditary connective tissue disorder testing, including single genes as well as multi-gene panels, which are defined as assays that simultaneously test for more than one hereditary connective tissue disorder gene. Medical necessity determination generally relies on criteria established for testing individual genes.

Medical necessity criteria differ based on the type of testing being performed (i.e., individual hereditary connective tissue disorder genes separately chosen versus pre-defined panels of genes) and how that testing will be billed (one or more individual gene procedure codes, specific panel procedure codes, or unlisted procedure codes).

Hereditary Connective Tissue Disorder single gene tests will be reimbursed when the following criteria are met:

The member has or is suspected to have a condition that will benefit from information provided by the requested hereditary connective tissue disorder gene testing based on at least one of the following:

The member displays clinical features of the condition for which testing is being requested and a genetic diagnosis would result in changes to the member’s medical management, OR

The member meets all criteria in a test-specific guideline, if available (see *table: Common hereditary connective tissue disorder genes, associated conditions, and applicable guidelines*), AND

The member does not have a known underlying cause for their symptoms (e.g. known genetic condition), AND

Rendering laboratory is a qualified provider of service per the Health Plan policy.

Hereditary Connective Tissue Disorder multi-gene panels will be reimbursed when the following criteria are met:

When separate procedure codes will be billed for individual hereditary connective tissue disorder genes (e.g., Tier 1 MoPath codes 81200-81355 or Tier 2 MoPath codes 81400-81408), each individually billed test will be evaluated separately. The following criteria will be applied:

The member has or is suspected to have a condition that will benefit from information provided by the requested hereditary connective tissue disorder gene testing based on at least one of the following:

The member displays clinical features of the condition for which testing is being requested and a genetic diagnosis would result in changes to the member's medical management, OR

The member meets all criteria in a test-specific guideline, if available, (see *Common hereditary connective tissue disorder genes, associated conditions, and applicable guidelines* table for a list of genes, associated conditions, and applicable guideline), AND

The member does not have a known underlying cause for their symptoms (e.g. known genetic condition), AND

Rendering laboratory is a qualified provider of service per the Health Plan policy.

When a patient meets medical necessity criteria for any hereditary connective tissue disorder gene(s) included in the panel, genetic testing for the clinically indicated gene(s) will be reimbursed. This includes the sequencing and deletion/duplication[†] components. Any genes that are included in a multi-gene panel but do NOT meet medical necessity criteria will NOT be a reimbursable service. It will be at the laboratory, provider, and patient's discretion to determine if a multi-gene panel remains the preferred testing option.

When a multi-gene panel is being requested and will be billed with a single panel CPT code (e.g. 81410, 81479) to represent all genes being sequenced, with or without another single procedure code representing the deletion/duplication[†] analysis portion, the panel will be considered medically necessary when the following criteria are met:

Medical necessity must be established for at least TWO conditions included in the panel based on the following:

The member displays clinical features of the condition for which testing is being requested and a genetic diagnosis would result in changes to the member's medical management, OR

The member meets all criteria in a test-specific guideline, if available, (see *table: Common hereditary connective tissue disorder genes, associated conditions, and applicable guidelines*), AND

The member does not have a known underlying cause for their symptoms (e.g. known genetic condition), AND

Clinical features are not sufficiently specific to suggest a single causative gene, AND

Rendering laboratory is a qualified provider of service per the Health Plan policy.

† When deletion/duplication testing is not part of a single panel CPT code being billed, deletion/duplication testing should be billed in only one of the following ways:

A separate CPT code for deletion/duplication analysis of each individual gene (may include non-specific molecular pathology tier 2 codes and/or unlisted code 81479), or

A single CPT code specific to the performed deletion/duplication analysis panel (e.g. 81411, 81479), or

A single microarray procedure (e.g. 81228 or 81229).

Procedure codes representing multiple methods for deletion/duplication testing will not be reimbursable for the same panel (e.g., test-specific deletion/duplication procedure codes and microarray will not both be reimbursable for the same panel).

Exceptions

The following are specifically non-reimbursed indications for Hereditary Connective Tissue Disorder testing:

Members personal and/or family history are suggestive of hypermobile EDS or the related clinical entity, "joint hypermobility syndrome"

Isolated joint hypermobility, including both asymptomatic and symptomatic forms (e.g., "hypermobility spectrum disorders")

Billing and Reimbursement Considerations

The billed amount should not exceed the list price of the test.

Broad connective tissue disorder panels may not be medically necessary when a narrower panel is available and more appropriate based on the clinical findings.

Genetic testing is only necessary once per lifetime. Therefore, a single gene included in a panel or a multi-gene panel may not be reimbursed if testing has been performed previously. Exceptions may be considered if technical advances in testing demonstrate significant advantages that would support a medical need to retest.

This guideline may not apply to genetic testing for indications that are addressed in test-specific guidelines. Please see the test-specific list of guidelines for a complete list of test-specific panel guidelines.

If a panel was previously performed and an updated, larger panel is being requested, only testing for the medically necessary, previously untested genes will be reimbursable. Therefore, only the most appropriate procedure codes for those additional genes will be considered for reimbursement.

Common Hereditary Connective Tissue Disorder Genes, Associated Conditions, and Applicable Guidelines

Common Hereditary Connective Tissue Disorder Genes, Associated Conditions, and Applicable Guidelines

<u>Condition</u>	<u>Gene</u>	<u>CPT</u>	<u>Applicable guideline</u>
<u>Arterial tortuosity syndrome</u>	<u>SLC2A10</u>	<u>81479</u>	<u>MOL.TS.268</u>
<u>Congenital contractural arachnodactyly</u>	<u>FBN2</u>	<u>81479</u>	<u>MOL.TS.268</u>
<u>Cutis laxa</u>	<u>ALDH18A1</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>ATP6V0A2</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>EFEMP2</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>ELN</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>FBLN5</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>LTBP4</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>PYCR1</u>	<u>81479</u>	<u>MOL.TS.268</u>

<u>Condition</u>	<u>Gene</u>	<u>CPT</u>	<u>Applicable guideline</u>
<u>Ehlers-Danlos syndrome (EDS)</u>	<u>ADAMTS2</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>B3GALT6</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>B4GALT7</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>C1R</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>C1S</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>CHST14</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>COL1A1</u>	<u>81408</u>	<u>MOL.TS.267</u>
	<u>COL1A2</u>	<u>81408</u>	<u>MOL.TS.267</u>
	<u>COL12A1</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>COL3A1</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>COL5A1</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>COL5A2</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>DSE</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>FKBP14</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>PLOD1</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>PRDM5</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>SLC39A13</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>TNXB</u>	<u>81479</u>	<u>MOL.TS.267</u>
	<u>ZNF469</u>	<u>81479</u>	<u>MOL.TS.267</u>

<u>Condition</u>	<u>Gene</u>	<u>CPT</u>	<u>Applicable guideline</u>
<u>FLNA deficiency (periventricular nodular heterotopia)</u>	<u>FLNA</u>	<u>81479</u>	<u>MOL.TS.268</u>
<u>Homocystinuria (cystathionine beta-synthase deficiency)</u>	<u>CBS</u>	<u>81401 81406</u>	<u>MOL.TS.268</u>
<u>Juvenile polyposis/hereditary hemorrhagic telangiectasia</u>	<u>SMAD4</u>	<u>81406</u>	<u>MOL.TS.268</u>
	<u>SMAD4</u>	<u>81405</u>	<u>MOL.TS.268</u>
<u>Loeys-Dietz syndrome</u>	<u>SMAD3</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>SMAD2</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>TGFB2</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>TGFB3</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>TGFBR1</u>	<u>81405</u>	<u>MOL.TS.268</u>
	<u>TGFBR2</u>	<u>81405</u>	<u>MOL.TS.268</u>
<u>MED12-related disorders</u>	<u>MED12</u>	<u>81401 81479</u>	<u>MOL.TS.268</u>
<u>Marfan syndrome</u>	<u>FBN1</u>	<u>81408</u>	<u>MOL.TS.202</u>
	<u>TGFBR1</u>	<u>81405</u>	<u>MOL.TS.202</u>
	<u>TGFBR2</u>	<u>81405</u>	<u>MOL.TS.202</u>
<u>NOTCH1-related aortic valve disease/Adams-Oliver syndrome</u>	<u>NOTCH1</u>	<u>81407</u>	<u>MOL.TS.268</u>
<u>Occipital horn syndrome/Menkes</u>	<u>ATP7A</u>	<u>81479</u>	<u>MOL.TS.268</u>

<u>Condition</u>	<u>Gene</u>	<u>CPT</u>	<u>Applicable guideline</u>
<u>Osteogenesis imperfecta</u>	<u>COL1A1</u>	<u>81408</u>	<u>MOL.TS.268</u>
	<u>COL1A2</u>	<u>81408</u>	<u>MOL.TS.268</u>
<u>Pseudoxanthoma elasticum</u>	<u>ABCC6</u>	<u>81479</u>	<u>MOL.TS.268</u>
<u>Shprintzen-Goldberg syndrome</u>	<u>SKI</u>	<u>81479</u>	<u>MOL.TS.268</u>
<u>Stickler syndrome</u>	<u>COL11A1</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>COL11A2</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>COL2A1</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>COL9A1</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>COL9A2</u>	<u>81479</u>	<u>MOL.TS.268</u>
	<u>COL9A3</u>	<u>81479</u>	<u>MOL.TS.268</u>
<u>Thoracic aortic aneurysm and dissection (TAAD)</u>	<u>ACTA2</u>	<u>81405</u>	<u>MOL.TS.227</u>
	<u>BGN</u>	<u>81479</u>	<u>MOL.TS.227</u>
	<u>COL3A1</u>	<u>81479</u>	<u>MOL.TS.227</u>
	<u>FBN1</u>	<u>81408</u>	<u>MOL.TS.227</u>
	<u>MAT2A</u>	<u>81479</u>	<u>MOL.TS.227</u>
	<u>MFAP5</u>	<u>81479</u>	<u>MOL.TS.227</u>
	<u>MYH11</u>	<u>81408</u>	<u>MOL.TS.227</u>
	<u>MYLK</u>	<u>81479</u>	<u>MOL.TS.227</u>
	<u>PRKG1</u>	<u>81479</u>	<u>MOL.TS.227</u>

<u>Condition</u>	<u>Gene</u>	<u>CPT</u>	<u>Applicable guideline</u>
	<u>SMAD3</u>	<u>81479</u>	<u>MOL.TS.227</u>
	<u>TGFB2</u>	<u>81479</u>	<u>MOL.TS.227</u>
	<u>TGFB3</u>	<u>81479</u>	<u>MOL.TS.227</u>
	<u>TGFBR1</u>	<u>81405</u>	<u>MOL.TS.227</u>
	<u>TGFBR2</u>	<u>81405</u>	<u>MOL.TS.227</u>

Note Several genes in this table are associated with multiple genetic disorders, including some not listed above. The test should be reviewed for the appropriate condition/indication.

References

Benedek TG, Rodnan GP. Connective Tissue Disease. (Updated September 2020). Encyclopedia Britannica. Available at: <https://www.britannica.com/science/connective-tissue-disease>

Connective Tissue Disorders (Last reviewed April 2016). In Medline Plus Health Topics. US National Library of Medicine. (database online). Copyright, National Institutes of health. 1993-2021. Available at: <https://medlineplus.gov/connectivetissuedisorders.html>

Callewaert B, De Paepe A, Coucke P. Arterial Tortuosity Syndrome. 2014 Nov 13 [Updated 2020 Nov 19]. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK253404/>.

Callewaert B. Congenital Contractural Arachnodactyly. 2001 Jan 23 [Updated 2019 Oct 21]. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022 Available at: <https://www.ncbi.nlm.nih.gov/books/NBK1386/>

Cutis Laxa. (Last reviewed July 2021). In: MedlinePlus Genetics US National Library of Medicine (database online). Copyright, National Institutes of Health. 1993-2022 Available at: <https://medlineplus.gov/genetics/condition/cutis-laxa/#genes>

Callewaert BL and Urban Z. LTBP4-Related Cutis Laxa. 2016 Feb 11 [Updated 2021 Jul 22]. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK343782/>.

Ehlers-Danlos Syndrome. (Last Reviewed August 2020). In: MedlinePlus Genetics US National Library of Medicine (database online). Copyright, National Institutes of Health. 1993-2020. Available at:

<https://medlineplus.gov/genetics/condition/ehlers-danlos-syndrome/>

Malfait F, Francomano C, Byers P, et al. The 2017 international classification of the Ehlers-Danlos syndromes. *Am J Med Genet C Semin Med Genet.* 2017 Mar;175(1):8-26.

Sacharow SJ, Picker JD, Levy HL. Homocystinuria Caused by Cystathionine Beta-Synthase Deficiency. 2004 Jan 15 [Updated 2017 May 18]. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022. Available at:

<https://www.ncbi.nlm.nih.gov/books/NBK1524/>

Loeys BL, Dietz HC. Loeys-Dietz Syndrome. 2008 Feb 28 [Updated 2018 Mar 1]. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022. Available at:

<https://www.ncbi.nlm.nih.gov/books/NBK1133/>

Dietz HC. FBN1-Related Marfan Syndrome. 2001 Apr 18 [Updated 2022 Feb 17]. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022. Available at:

<https://www.ncbi.nlm.nih.gov/books/NBK1335/>

Lehman A, Wuyts W, Patel MS. Adams-Oliver Syndrome. 2016 Apr 4. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022. Available at:

<https://www.ncbi.nlm.nih.gov/books/NBK355754/>

Steiner RD, Basel D. COL1A1/2-Related Osteogenesis Imperfecta. 2005 Jan 28 [Updated 2021 May 6]. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK1295/>.

Chen MH, Walsh CA. FLNA Deficiency. 2002 Oct 8 [Updated 2021 Sep 30]. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022. Available at:

<https://www.ncbi.nlm.nih.gov/books/NBK1213/>

Robin NH, Moran RT, Ala-Kokko L. Stickler Syndrome. 2000 Jun 9 [Updated 2021 May 6]. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022. Available at:

<https://www.ncbi.nlm.nih.gov/books/NBK1302/>

Milewicz DM, Regalado E. Heritable Thoracic Aortic Disease Overview. 2003 Feb 13 [Updated 2017 Dec 14]. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK1120/>.

Levy, HP. Hypermobile Ehlers-Danlos syndrome. 2004 Oct 22 [Updated 2018 Jun 21]. In: Adam MP, Ardinger HH, Pagon RA, et al., editors. GeneReviews® [Internet]. Seattle (WA): University of Washington, Seattle; 1993-2022. Available at: <https://www.ncbi.nlm.nih.gov/books/NBK1279/O.20.00046>